Innovation

A new diagnostic test for Creutzfeldt-Jakob disease: Real-time quakinginduced conversion (RT-QulC)

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Abstract

Currently, definitive diagnoses of Creutzfeldt-Jakob disease (CJD) require an examination of the brains of deceased patients. In cooperation with an international consortium, Canada's National Microbiology Laboratory is performing a validation of a new test that can diagnose CJD in living patients. This test, the real-time quaking-induced conversion (RT-QuIC) assay, directly detects the disease associated form of the prion protein in cerebrospinal fluid. If validated, RT-QuIC will represent a major advance in the diagnoses of patients suspected to have CJD.

Introduction

Creutzfeldt-Jakob disease (CJD) is a rare, untreatable, uniformly fatal brain disorder. The disease most commonly affects older adults at a rate of one case of CJD diagnosed per million people each year (1). The first signs often include anxiety, confusion and memory loss resembling those of other neurological disorders such as Alzheimer's disease and Huntington's disease. There are several subtypes of CJD and the signs of disease progression are highly variable. However, it is common to observe difficulties with balance, hand coordination, walking, vision and swallowing, as well as involuntary jerky movements (2). Ultimately, people lose their ability to move and speak; death is often caused by pneumonia or other secondary infections.

These fatal neurological diseases are caused by novel transmissible agents called prions that are not conventional viruses or bacteria but rather are composed of an abnormal form of a host protein. This key biological fact precludes the use of technologies currently applied to the direct detection of other infectious agents. Presently, definitive diagnoses of CJD require an examination of the brains of deceased patients revealing prion protein deposits and associated nerve cell loss resulting in the brain having a sponge-like appearance.

The objective of this article is to summarize three different types of CJD, to describe a new method recently developed to diagnose CJD, and to consider the potential implications for patients and clinical/laboratory practice.

Classification of CJD

Creutzfeldt-Jakob disease can be classified into three broad categories: sporadic CJD, genetic CJD and acquired CJD. Sporadic CJD (sCJD) represents up to 85% of observed cases, develops for no apparent reason and usually affects people in their 60s. Depression is a common early symptom and most people with sCJD die within six months of diagnosis.

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Genetic CJD (gCJD) can present like sporadic CJD or may present as a slowly progressing dementia that develops over years. Some of the more common CJD mutations have been tracked through large multigenerational families and display autosomal dominant inheritance. Nonetheless, in 50% of genetic CJD diagnoses there is no familial history of CJD or other neurological disorders suggesting low penetrance (3).

Acquired CJD, accounting for less than 1% of cases, results from exposure to the infectious agent. Iatrogenic CJD describes cases of acquired CJD related to medical procedures. New variant CJD (vCJD) is acquired through dietary exposure to beef infected with bovine spongiform encephalopathy (BSE), the scientific term for mad cow disease. Due to the effective implementation of public health measures (4), the risk of acquiring variant CJD in Canada is nearly nonexistent. Indeed, there has never been a case of domestically acquired variant CJD reported in Canada (1).

Current practice

Pre-mortem diagnoses of any form of CJD rely on clinical presentation, characteristic electroencephalogram (EEG) and magnetic resonance imaging (MRI) patterns. The appearance of surrogate biomarkers, typically a change in the amount of specific proteins in cerebrospinal fluid (CSF), provides additional supporting diagnostic data.

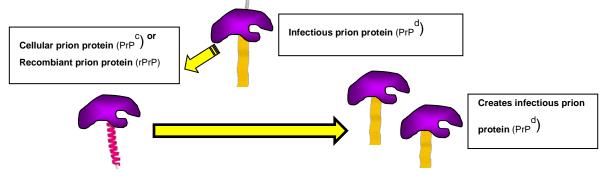
Currently, the Prion Laboratory Section at the National Microbiology Laboratory measures the level of three proteins (14-3-3, tau and S100) in CSF, to support clinicians dealing with suspected cases of CJD. Increased levels of these three surrogate markers are consistent with a diagnosis of CJD, but they have variable sensitivity and specificity and consequently cannot be used to definitively diagnose CJD (5).

The molecular basis of the new test

At the molecular level CJD disease progression is characterized by the conversion of the normally expressed cellular prion protein, PrP^c , into the misfolded disease associated isoform, PrP^d (**Figure 1**). The infectious form of the prion protein, PrP^d , acts as a "seed" and interacts with normal cellular form of the prion protein, PrP^c , and promotes its conversion to the infectious form. As the disease progresses the amount of PrP^d available to convert PrP^c increases, accelerating the conversion of PrP^d . Eventually this leads to the generation of the large amounts of PrP^d observed in the brains of CJD patients post-mortem.

In contrast to what is observed in the brain, only low amounts of PrP^d are present in other tissues making detection in an easily accessible sample problematic. A solution to this challenge has been the development of a technique, known as real-time quaking-induced conversion (RT-QuIC). RT-QuIC exploits the ability of PrP^d to promote the conformational conversion of PrP^c (6-8). In RT-QuIC a recombinant prion protein (rPrP) made in the laboratory is used as the substrate and patient CSF samples, potentially containing PrP^d, are tested to see if they will "seed" the conversion process (**Figure 1**). Cycles of shaking and incubation will cause infected CSF, containing PrP^d, to generate greatly increased amounts of rPrP^d facilitating subsequent detection by traditional methods.

Figure 1: Infectious prion protein promptes the conformational conversion of cellular prion protein



Legend: In RT-QuIC the PrP^d present in infected samples interacts with laboratory produced rPrP causing a conformational conversion that can be monitored in real time.

Summary of test results

RT-QuIC has been demonstrated to be a highly sensitive and specific method to detect the presence of the disease-associated isoform of the prion protein, PrP^d, in the CSF of patients with sporadic CJD (9-13). Direct detection of the disease-causing agent is a major advance in the development of a diagnostic test for sporadic CJD as it can be used for definitive diagnoses while the patient is still alive. However, ongoing validation studies have demonstrated that the outcomes of RT-QuIC reactions depend on the exact conditions employed (biochemical, chemical and physical), as well as the nature of the sample and recombinant substrate (14). The protocol under evaluation by the National Microbiology Laboratory has been tested extensively for the detection of sporadic CJD (10,11,15,16). Using this protocol the Prion Laboratory Section achieved 100% sensitivity and 100% specificity in the evaluation of an external sporadic CJD infected CSF proficiency panel. This is comparable to the 91% sensitivity and 98% specificity reported by other laboratories (8,9). Being part of an international consortium using the same standardized protocol should permit analyses of sufficient numbers of the rarer vCJD and gCJD samples to determine the efficacy of our version of RT-QuIC in these cases.

Discussion

The establishment of a specific and sensitive test will aid health care workers as well as patients and their families in dealing with suspected cases of CJD. A more specific and sensitive diagnostic test for CJD will also help address wider public health considerations concerning the transmission of CJD from patient to patient through the use of contaminated surgical instruments and tissues, including blood. Once validation is completed RT-QuIC will be a service offered by the Prion Laboratory Section of the National Microbiology Laboratory, making a more sensitive and specific ante-mortem test for CJD available to Canadian health care professionals.

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Conflict of interest

None

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References

- (1) Coulthart MB, Jansen GH, Connolly T, D'Amour R, Kruse J, Lynch J et al. Creutzfeldt-Jakob disease mortality in Canada, 1998–2013 Can Comm Dis Rep 2015;41:182-191.
- (2) Rosenbloom MH, Atri A. The evaluation of rapidly progressive dementia. Neurologist. 2011;17(2):67–74.
- (3) Kovács GG, Puopolo M, Ladogana A, Pocchiari M, Budka H, van Duijn C, Collins SJ, Boyd A, Giulivi A, Coulthart M, Delasnerie-Laupretre N, Brandel JP, Zerr I, Kretzschmar HA, de Pedro-Cuesta J, Calero-Lara M, Glatzel M, Aguzzi A, Bishop M, Knight R, Belay G, Will R, Mitrova E. Genetic prion disease: The EUROCJD experience. Hum Genet. 2005 Nov;118(2):166–74.
- (4) Canadian Food Inspection Agency. Evaluation of Bovine Spongiform Encephalopathy (BSE) Management Program. http://www.inspection.gc.ca/about-the-cfia/accountability/other-activities/audits-reviews-and-evaluations/evaluation-of-the-bse-management-program/evaluation/eng/1419265462091/1419265463216
- (5) Coulthart MB, Jansen GH, Olsen E, Godal DL, Connolly T, Choi BC, Wang Z, Cashman NR. Diagnostic accuracy of cerebrospinal fluid protein markers for sporadic Creutzfeldt-Jakob disease in Canada: A 6-year prospective study. BMC Neurol. 2011 Oct 27;11:133. doi: 10.1186/1471-2377-11-133.
- (6) Saborio GP, Permanne B, Soto C. Sensitive detection of pathological prion protein by cyclic amplification of protein misfolding. Nature. 2001;411(6839):810−3.
- (7) Atarashi R, Moore RA, Sim VL, Hughson AB, Dorward D, Onwubiko HA, Priola SA, Caughey B. Ulrtasensitive detection of scrapie prion protein using seeded conversion of recombinant prion protein. Nat Methods. 2007;4(8):645–50.
- (8) Wilham JM, Orrú CD, Bessen RA, Atarashi R, Sano K, Race B, Meade-White KD, Taubner LM, Timmes A, Caughey B. Rapid end-point quantitation of prion seeding activity with sensitivity comparable to bioassays. PLoS Pathog. 2010;Dec 2;6(12):e1001217.
- (9) Atarashi R, Satoh K, Sano K, Fuse T, Yamaguchi N, Ishibashi D, Matsubara T, Nakagaki T, Yamanaka H, Shirabe S, Yamada M, Mizusawa H, Kitamoto T, Klug G, McGlade A, Collins SJ, Nishida N. Ultrasensitive human prion detection in cerebrospinal fluid by real-time quaking-induced conversion. Nat. Med. 2011;17:175–8.
- (10) Peden AH, McGuire LI, Appleford NEJ, Mallinson G, Wilham JM, Orrú CD, Caughey B, Ironside JW, Knight RS, Will RG, Green AJE, Head MW. Sensitive and specific detection of sporadic Creutzfeldt–Jakob disease brain prion protein using real-time quaking-induced conversion. J. Gen Virol. 2012;93:438–49.
- (11) McGuire LI, Peden AH, Orrú CD, Wilham JM, Appleford NE, Mallinson G, Andrews M, Head MW, Caughey B, Will RG, Knight RS, Green AJ. Real time quaking-induced conversion analysis of cerebrospinal fluid in sporadic Creutzfeldt-Jakob disease. Ann Neurol. 2012;72:278–85.
- (12) Cramm M, Schmitz M, Karch A, Zafar S, Varges D, Mitrova E, Schroeder B, Raeber A, Kuhn F, Zerr I. Characteristic CSF prion seeding efficiency in humans with prion diseases. Mol Neurobiol. 2015 Feb;51(1):396–405. doi: 10.1007/s12035-014-8709-6.
- (13) Orrú CD, Bongianni M, Tonoli G, Ferrari S, Hughson AG, Groveman BR, Fiorini M, Pocchiari M, Monaco S, Caughey B, Zanusso G. A test for Creutzfeldt-Jakob disease using nasal brushings. N Engl J Med. 2014;371:519–29.
- (14) Cheng K, Sloan A, Avery KM, Coulthart M, Carpenter M, Knox JD. Exploring physical and chemical factors influencing the properties of recombinant prion protein and the real-time quaking-induced conversion (RT-QuIC) Assay. PloS One. 2014 Jan3;9:1. doi: 10.1371/journal.pone.0084812.
- (15) Orrú CD, Wilham IM. Raymond LD, Kuhn F, Schroeder B, Raiber AI, Caughey B. Prion disease blood test using immunoprecipitation and improved quaking-induced conversion. mBio. 2011;2e00078-11.
- (16) Sano K, Satoh K, Atarashi R, Takashima H, Iwasaki Y, Yoshida M, Sanjo N, Murai H, Mizusawa H, Schmitz M, Zerr I, Kim YS, Nishida N. Early detection of abnormal prion protein in genetic human prion diseases now possible using real-time QUIC assay. PLoS One. 2013; 8(1):e54915.